

Instructions for Use

Version: 2.1.1

Revision date: 29-Aug-25



Albumin Assay Kit

Catalog No.: abx298891

Size: 96 tests

Detection Range: 0.08 g/L – 15 g/L

Sensitivity: 0.08 g/L

Storage: Store the Chromogenic Reagent at 4°C in the dark. Store the Standards at -20°C.

Application: For detection and quantification of Albumin concentration in serum, plasma, and cell culture supernatants.

Introduction

Abbexa's Albumin Assay Kit is a quick, convenient, and sensitive method for measuring and calculating Albumin concentration. Bromocresol Green dye combines with Albumin at pH 4.0 – 4.2, forming a blue-green colored complex with an absorbance maximum at 630 nm. The intensity of the color is proportional to the Albumin concentration, which can then be calculated.

Kit components

1. 96-well microplate
2. Chromogenic Reagent: 6 ml
3. Standard (20 g/L): 2 × 1.2 ml
4. Plate sealer: 2

Materials required but not provided

1. Microplate reader (630 nm)
2. Double-distilled water
3. Normal saline (0.9 % NaCl)
4. PBS (0.01 M, pH 7.4)
5. Pipette and pipette tips
6. 1.5 ml microcentrifuge tubes
7. Vortex mixer
8. Incubator

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Protocol

A. Preparation of samples and reagents

1. Reagents

- **Chromogenic Reagent Working Solution:** Prepare enough Chromogenic Reagent Working Solution for the wells tested. Mix the Chromogenic Reagent and double-distilled water in a ratio of 1 : 4. Prepare 250 µl Chromogenic Reagent Working Solution per well by mixing 50 µl Chromogenic Reagent with 200 µl double-distilled water. Prepare immediately before use.
- **Standards:** Standard vials should be taken from storage and placed on ice to thaw gradually. Label 7 tubes with 1.0 g/L, 2.0 g/L, 3.5 g/L, 5.0 g/L, 8.0 g/L, 12.0 g/L, and 15.0 g/L. Prepare these dilutions according to the volumes in the following table:

Standard Dilution (g/L)	1.0	2.0	3.5	5.0	8.0	12.0	15.0
20 g/L Standard (µl)	10	20	35	50	80	120	150
Double distilled water (µl)	190	180	165	150	120	80	50

For the blank, use pure double-distilled water. The volume of each standard dilution will be 200 µl.

Note:

- Allow all Chromogenic Reagent to equilibrate to room temperature before use.
- Keep Standard on ice during use, avoid repeated freeze-thaw cycles.

2. Samples

Isolate the test samples soon after collecting and analyze immediately or aliquot and store at -80°C for up to one month. Avoid multiple freeze-thaw cycles.

- **Serum/Plasma:** Samples can be tested directly.
- **Cell Culture Supernatant:** Samples can be tested directly.

It is recommended to carry out a preliminary experiment to determine the optimal dilution factor of samples before carrying out the formal experiment. Dilute samples into different concentrations with normal saline (0.9 % NaCl) or PBS (0.01 M, pH 7.4), then carry out the assay procedure. The recommended dilution factor for different samples is as follows (for reference only):

Sample Type	Dilution Factor
Human serum	8 – 15
Human plasma	8 – 15
HepG2 supernatant	1
Mouse plasma	8 – 15
Rat serum	8 – 15

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Note:

- Fresh samples or recently obtained samples are recommended to prevent degradation and denaturalization that may lead to erroneous results.

B. Assay Procedure

- Mark positions on the 96-well microplate for each standard, blank, and sample. *It is strongly recommended to test all wells in duplicate. Avoid foaming or bubbles.*
- Add 10 µl of each standard dilution to the corresponding standard wells.
- Add 10 µl of sample each sample well.
- Add 250 µl Chromogenic Reagent Working Solution to each well.
- Incubate for 10 minutes at room temperature.
- Measure the OD of each well with a microplate reader at 630 nm.

C. Calculation of Results

Plot the standard curve, using the mean OD of the standard dilutions (adjusted for the blank) on the y-axis, and their respective concentrations on the x-axis. The curve should form a straight line, described by the formula $y = ax + b$.

Based on this curve, the concentration of Albumin in each sample well can be derived using the following formula:

1. Serum/Plasma/Cell Culture Supernatant samples:

$$\text{Albumin Concentration (g/L)} = \frac{(\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}} - b)}{a} \times F$$

where:

OD _{Sample}	OD of the sample wells
OD _{Blank}	OD of the blank wells
a	Gradient of the standard curve ($y = ax + b$)
b	Y-intercept of the standard curve ($y = ax + b$)
F	Dilution factor of the sample

Technical Support

For troubleshooting and technical assistance, please contact us at support@abbexa.com.