

# Instructions for Use

Version: 2.0.0

Revision date: 04 Jan 2023



## Human Glycated Hemoglobin (HbA1c) CLIA Kit

**Catalog No:** abx490036

**Size:** 96T

**Range:** 5.86 µg/ml - 1500 µg/ml

**Sensitivity:** < 2.44 µg/ml

**Storage:** Store the 96-well plate, Standards, and Detection Reagent(s) at -20°C, and the rest of the kit components at 4°C.

**Application:** The quantitative detection of HbA1c in Human Erythrocyte lysates, serum and plasma.

**Principle of the Assay:** This kit is based on competitive chemiluminescent immunoassay technology. An antibody is pre-coated onto a 96-well plate. Standards, test samples, and biotin-conjugated reagent are added to the wells and incubated. A competitive inhibition reaction takes place between the biotin-labelled HbA1c and the unlabelled-HbA1c on the pre-coated antibody. The HRP-conjugated reagent is then added, and the whole plate is incubated. Unbound conjugates are removed using wash buffer at each stage. After the Substrate working solution is added, only wells that contain the labelled HbA1c will produce chemiluminescence. The intensity of the emitted light is inversely proportional to the amount of HbA1c in the sample.

### Kit Components

- Pre-coated 96-Well Microplate: 12 x 8
- Standard: 2 tubes
- Wash Buffer: (30X) 20 ml
- Standard Diluent Buffer: 20 ml
- Detection Reagent A: (100X) 120 µl
- Detection Reagent B: (100X) 120 µl
- Diluent A: 12 ml
- Diluent B: 12 ml
- Substrate A: 10 ml
- Substrate B: 2 ml
- Plate Sealer: 3

### Materials Required But Not Provided

- 37°C incubator
- Multi and single channel pipettes and sterile pipette tips
- Squir bottle or automated microplate washer
- 1.5 ml tubes
- Distilled water
- Absorbent filter papers
- 100 ml and 1 liter graduated cylinders
- 0.01 mol/L PBS (pH 7.0 - 7.2)
- Luminometer capable of reading 96-well microplates (lag time 30.0 secs and read time 1.0 sec/well)

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## Protocol

### A. Sample Preparation

Analyse immediately or store samples at 2-8°C (within 24 hrs). For long term storage, aliquot and store at -20°C or -80°C. Avoid multiple freeze-thaw cycles.

- **Serum:** Samples should be collected into a serum separator tube. Coagulate the serum by leaving the tube undisturbed in a vertical position overnight at 4°C or at room temperature for up to 1 hr. Centrifuge at approximately 1000 × g for 20 mins. If precipitate appears, centrifuge again. Assay immediately or aliquot and store at -20°C or -80°C.
- **Erythrocyte lysates:** Centrifuge whole blood for 20 minutes at 1000 x g. Discard the supernatant and collect the pellet. Wash the cells three times in ice cold PBS, then resuspend cells in ice cold PBS. Freeze the cells at -20C, then thaw by gentle mixing at 4C. Repeat the freeze-thaw cycle three times. Centrifuge for 10 minutes at 5000 x g at 4C. Collect the supernatant and assay immediately, or aliquot and store at 4C for up to 5 days, 20C for up to 1 month or 80C for up to 2 months. Avoid repeated freeze-thaw cycles. Note: erythrocyte lysate samples likely require dilution before assay.
- **Plasma:** Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 mins at 1000 x g, within 30 mins of collection. If precipitate appears, centrifuge again. Avoid hemolytic samples.

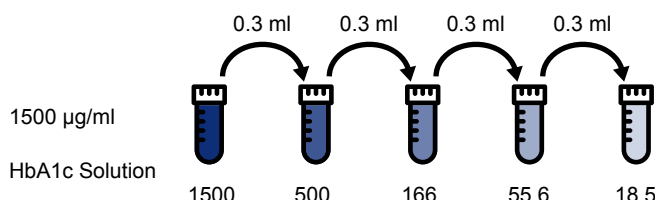
### Notes:

- Samples must be diluted so that the expected concentration falls within the kit's range. The sample should be diluted in 0.01 mol/L PBS (PH=7.0-7.2).
- Always use non-pyrogenic, endotoxin-free tubes for blood collection.
- Fresh samples, or recently obtained samples, are recommended to prevent protein degradation and denaturation that may lead to erroneous results.
- NaN<sub>3</sub> cannot be used as a test sample preservative, since it inhibits HRP.
- If possible, prepare solid samples using sonication and/or homogenization, as lysis buffers may (on occasion) interfere with the kit's performance.
- If a sample is not indicated in the manuals applications, a preliminary experiment to determine the suitability of the kit will be required.

### B. Reagent Preparation

**Standard:** Prepare the Standard with 1 ml of Standard Diluent buffer to make the 1500 µg/ml Standard Solution. This is the highest standard. Allow the reconstituted standard to sit for 10 mins, with gentle agitation prior to carrying out the serial dilutions. Avoid foaming or bubbles. Label tubes in preparation for the serial dilutions - see the diagram below for reference. Aliquot 0.6 ml of the Standard Diluent Buffer into each tube (apart from the highest standard tube). Add 0.3 ml of the highest standard solution into the 1st tube and mix thoroughly. Transfer 0.3 ml from the 1st to 2nd tube, mix thoroughly, and so on.

**Note:** Do not vortex the standard during reconstitution, as this will destabilize the protein. Once the standard has been reconstituted, it should be used within 15 mins. It is not recommended to reuse the reconstituted standard.



**Wash Buffer:** Dilute the concentrated Wash Buffer 30-fold (1/30) with distilled water (i.e. add 20 ml of concentrated wash buffer into 580 ml of distilled water). If crystals have formed in the concentrated Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.

**Detection Reagent A Working Solution Preparation:** Prepare no more than 15 mins before the experiment.

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1. Calculate the total volume of working solution required.
2. Dilute Detection Reagent A 100-fold with Diluent A, and mix thoroughly. Pipette with a slow, smooth action to reduce volume errors.

**Detection Reagent B Working Solution Preparation:** Prepare no more than 15 mins before the experiment.

1. Calculate the total volume of working solution required.
2. Dilute Detection Reagent B 100-fold with Diluent B, and mix thoroughly. Pipette with a slow, smooth action to reduce volume errors.

**Preparation of Substrate Working Solution:**

1. Calculate the total volume of working solution required.
2. In a separate tube, add Substrate A and Substrate B in a 99:1 ratio, respectively (e.g. add 990  $\mu$ l Substrate A to 10  $\mu$ l Substrate B). Prepare no more than 10 mins before use.

## C. Assay Protocol

Prepare all standards, samples and reagents as directed above. Equilibrate the kit components and samples to room temperature prior to use. It is recommended to measure in duplicate, and to plot a standard curve for each test.

1. Set standard, test sample and control (zero) wells on the pre-coated plate respectively, and record their positions. *Add the solution to the bottom of each well without touching the side walls. Pipette the standards and samples up and down to mix before adding to the wells. Avoid foaming or bubbles.*
2. Aliquot 50  $\mu$ l of the diluted standards into the standard wells.
3. Aliquot 50  $\mu$ l of Standard Diluent buffer into the control (zero) well.
4. Aliquot 50  $\mu$ l of appropriately diluted sample into the test sample wells. Gently tap the plate to mix, or use a microplate shaker.
5. Immediately aliquot 50  $\mu$ l of Detection Reagent A working solution to each well. Gently tap the plate to mix, or use a microplate shaker. Cover the plate with a plate sealer and incubate for 1 hr at 37°C.
6. Remove the cover and discard the solution. Wash the plate 3 times with 1X Wash Buffer. *Fill each well completely with Wash buffer (350  $\mu$ l) using a multi-channel Pipette or autowasher (1-2 mins soaking period is recommended). Complete removal of liquid at each step is essential for good performance. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean absorbent paper towels.*
7. Aliquot 100  $\mu$ l of Detection Reagent B working solution to each well. Seal the plate and incubate for 30 mins at 37°C.
8. Remove the cover, discard the solution and repeat the wash process as described above, 5 times.
9. Aliquot 100  $\mu$ l of Substrate working solution into each well. Cover the plate with a plate sealer and incubate for 10 mins at 37°C.
10. Measure the chemiluminescence signal in a microplate luminometer immediately.

This assay is competitive, therefore there is an inverse correlation between the concentration of the sample and the RLUs (Relative Light Units) measured. Create a graph with the log of the standard concentration (Y) and OD measured (X). Apply a best fit trendline through the standard points. Use this graph to calculate sample concentrations based on their OD values. If the samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

### Precautions:

- Before using the kit, centrifuge the tubes to bring down the contents trapped in the lid.
- Do not leave the wells uncovered for extended periods between incubations. The addition of reagents for each step should not exceed 10 mins.
- Ensure that the plate is properly sealed or covered during the incubation steps, and that the time and temperature are controlled.
- Do not reuse pipette tips and tubes.
- Do not use expired components, or components from a different kit.
- Please note that this kit is optimised for detection of native samples, rather than recombinant proteins or synthetic chemicals. We are unable to guarantee detection of recombinant proteins, as they may have different sequences or tertiary structures to the native protein.

### Precision:

Intra-assay Precision (Precision within an assay): 3 samples with low, medium and high levels of HbA1c were tested 20 times on one plate, respectively.

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Inter-assay Precision (Precision between assays): 3 samples with low, medium and high levels of HbA1c were tested on 3 different plates, 8 replicates in each plate.

CV (%) = (Standard Deviation / Mean) × 100

Intra-Assay: CV < 10%

Inter-Assay: CV < 12%

For Reference Only