

## Prion Protein (PRNP) Antibody Pair

Catalogue No.:abx370619

Prion Protein (PRNP) Antibody Pair for use in Sandwich ELISA assay development. This antibody pair contains:

Component	5 × 96 tests	10 × 96 tests
Capture Antibody	200 µg	400 µg
Biotin-Conjugated Detection Antibody	50 µg	100 µg
Standard	2 µg	10 µg

Please note that quantities and concentrations may change between different batches.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target:	Prion Protein (PRNP)
Reactivity:	Rat
Tested Applications:	ELISA
Recommended dilutions:	Dilute the Capture Antibody 125-fold with Coating Buffer. Dilute the Biotin-Conjugated Detection Antibody 200-fold with Detection Antibody Diluent. Optimal dilutions/concentrations should be determined by the end user.
Form:	Liquid (Capture Antibody and Detection Antibody)
Reconstitution:	Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.
Storage:	Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles.
Buffer:	The Capture and Detection Antibody both contain 0.1% sodium azide.
Standard Form:	Lyophilized
Assay Type:	Sandwich
Capture Antibody Conjugation:	Unconjugated
Detection Antibody Conjugation:	Biotin
Concentration:	Capture Antibody: 0.5 mg/ml Biotin-Conjugated Detection Antibody: 0.2 mg/ml

Note:THIS PRODUCT IS FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC,THERAPEUTIC OR COSMETIC PROCEDURES. NOT FOR HUMAN OR ANIMAL
CONSUMPTION.
<ul> <li>Directions for use:</li> <li>Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vial before use. Working solutions should be prepared and used immediately. <u>Recommended Procedure:</u></li> <li>1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4°C overnight or at 37 °C for 2 hours</li> <li>2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.</li> <li>3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1,5 hours.</li> <li>4. Repeat the aspiration/wash process in Step 2.</li> <li>5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.</li> <li>6. Repeat the aspiration/wash process in Step 2.</li> <li>7. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.</li> <li>8. Repeat the aspiration/wash process in Step 2.</li> <li>9. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10 µl.</li> <li>10. Repeat the aspiration/wash process in Step 2.</li> <li>11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10 µl.</li> <li>12. Add Substrate Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing 13. Measure the absorbance immediately using a microplate reader set at 450 nm.</li> </ul>