Datasheet

Version: 2.0.0 Revision date: 10 Jun 2025



Immunoglobulin G (IgG) Antibody Pair

Catalogue No.:abx370730

Immunoglobulin G (IgG) Antibody Pair for use in Sandwich ELISA assay development. This antibody pair contains:

96 tests	10 × 96 tests
	96 tests

Capture Antibody 200 μg 400 μg Biotin-Conjugated Detection Antibody 50 μg 100 μg Standard 2 μg 10 μg

Please note that quantities and concentrations may change between different batches.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target: Immunoglobulin G (IgG)

Reactivity: Sheep

Tested Applications: ELISA

Recommended dilutions: Dilute the Capture Antibody 125-fold with Coating Buffer.

Dilute the Biotin-Conjugated Detection Antibody 200-fold with Detection Antibody Diluent.

Optimal dilutions/concentrations should be determined by the end user.

Form: Liquid (Capture Antibody and Detection Antibody)

Reconstitute the standard with Standard Diluent. The volume, and therefore standard

concentration, should be determined by the end user.

Storage: Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles.

Buffer: The Capture and Detection Antibody both contain 0.1% sodium azide.

Standard Form: Lyophilized

Assay Type: Sandwich

Capture Antibody Conjugation: Unconjugated

Detection Antibody Biotin

Conjugation:

Concentration: Capture Antibody: 0.5 mg/ml

Biotin-Conjugated Detection Antibody: 0.2 mg/ml

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Note:

THIS PRODUCT IS FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC, THERAPEUTIC OR COSMETIC PROCEDURES. NOT FOR HUMAN OR ANIMAL CONSUMPTION.

Directions for use:

Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

- 1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
- 2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
- 3. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
- 4. Repeat the aspiration/wash process in Step 2.
- 5. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
- 6. Repeat the aspiration/wash process in Step 2.
- 7. Add appropriately diluted Biotin-Conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
- 8. Repeat the aspiration/wash process in Step 2.
- 9. Add appropriately diluted Streptavidin HRP (100 μl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
- 10. Repeat the aspiration/wash process in Step 2.
- 11. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
- 12. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
- 13. Measure the absorbance immediately using a microplate reader set at 450 nm.

