Datasheet

Version: 3.0.0 Revision date: 09 Oct 2025



DNA Polymerase Mix

Catalogue No.:abx461002

Abbexa's DNA Polymerase Mix is a convenient ready-to-go 2X reaction mix designed to maximize experiment reproducibility. It contains DNA Polymerase (abx461001), MgCl₂ and ultra-pure dNTPs. The mix is optimized and ready-to-use, simply add water, template and primers. DNA Polymerase Mix (2X) dramatically reduces the time needed to set up reactions, thereby minimizing the risk of contamination. Greater reproducibility is ensured, by a reduction in the number of pipetting steps that can lead to pipetting errors. The 500 rxns size is provided as 10 × 1.25 ml DNA Polymerase Mix, and 1.2 ml 50 mM MgCl₂ solution for optional optimization of reaction conditions.

Target: DNA Polymerase Mix

Tested Applications: PCR

Form: Liquid

Biological Activity: One unit is defined as the amount of enzyme that incorporates 10 nmol of dNTPs into acid-insoluble

form in 30 minutes at 72 °C.

Storage: Store all components at -20°C. It is not recommended to store the enzyme at -80°C as ice crystals may

form on the active site, which can affect the enzyme activity. Avoid repeated freeze/thaw cycles.

Validity: Up to 12 months.

Buffer: The DNA Polymerase Mix (2X) contains 4 mM Mg²⁺.

Concentration: 2X

Note: THIS PRODUCT IS FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC, THERAPEUTIC

OR COSMETIC PROCEDURES. NOT FOR HUMAN OR ANIMAL CONSUMPTION.

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Directions for use: Reaction Components:

Component Volume

Template Variable, as required

Primers (20 μM each) 1 μl DNA Polymerase Mix (2X)25 μl

Water Variable, up to 50 µl

Total Volume 50 μl Thermal Cycling Conditions:

Step	Number of CyclesTemperature		Time per Cycle
Initial Denaturation1 cycle		95-98 °C	3 min
Denaturation		95-98 °C	15 seconds
Annealing	25-35 cycles	55-60 °C (primer depende	ent) 15 seconds
Extension		72 °C	1.5-2 kb/min
Notes:			

- The 2X mix contains 4 mM Mg²⁺ (final concentration 2 mM Mg²⁺), which is sufficient for amplification of most targets and should only be adjusted if necessary.
- Forward and reverse primers are generally used at a final concentration of 0.2-0.6 μ M each. It is recommended to start with 0.4 μ M as the final concentration (i.e. 20 pmol of each primer per 50 μ l reaction volume). A primer concentration that is too high can reduce the specificity of priming, resulting in non-specific products. Primers should have a melting temperature (T_m) of approximately 60 °C.
- The amount of template in the reaction depends mainly on the type of DNA used. For templates with low structural complexity, such as plasmid DNA, it is recommended to use 50 pg-10 ng DNA per 50 µl reaction volume. For eukaryotic genomic DNA, it is recommended to use a starting amount of 200 ng DNA per 50 µl reaction, this can be varied between 5 ng 500 ng. It is important to avoid using templates re-suspended in EDTA-containing solutions (e.g. TE buffer) since EDTA chelates free Mg²⁺.