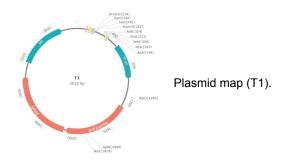


# **T1 Cloning Kit**

Catalogue No.:abx098055



T1 Cloning Kit is designed for cloning and sequencing Taq amplified PCR products. 5 minutes fast ligation of Taq-amplified PCR products. T7 promoter primer, M13 forward primer and M13 reverse primer for sequencing. T7 promoter for in vitro transcription.

# Kit contents:

| Component   | 20 rxns    | 60 rxns             |
|---|------------|---------------------|
| T1 Cloning Vector (10 ng/μl)                              | 20 µl      | $3 \times 20 \mu$ l |
| Control Template (5 ng/µl)                                | 5 µl       | 5 µl                |
| Control Primer (10 µM)                                    | 5 µl       | 5 µl                |
| M13 Forward Primer (10 µM)                                | 50 µl      | 150 µl              |
| M13 Reverse Primer (10 µM)                                | 50 µl      | 150 µl              |
| T1 Phage Resistant Chemically Competent Cells (abx098070) | 10 × 100 I | الا 100 × 100 ال    |

Target: T1 Cloning Kit

Storage: Store Phage Resistant Chemically Competent Cells at -70°C for up to 6 months and all other components at -20°C for up to 9 months.

Note: THIS PRODUCT IS FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC, THERAPEUTIC OR COSMETIC PROCEDURES.

NOT FOR HUMAN OR ANIMAL CONSUMPTION.

This product is shipped with dry ice.

# Datasheet

Version: 7.0.0 Revision date: 21 Oct 2025



#### **Directions for**

#### Preparation

use:

- 1. Add 0.5  $\mu$ l 4  $\mu$ l of PCR products and 1  $\mu$ l of T1 Cloning Vector into a microcentrifuge tube.
- 2. Gently mix, incubate at room temperature for 5 minutes, and place the tube on ice.

The optimal molar ratio of the vector to insert is 1:7 (for example, 1 kb = 20 ng, 2 kb = 40 ng). The optimal vector volume is 1  $\mu$ l. The total reaction volume should be 3  $\mu$ l - 5  $\mu$ l. The optimal incubation temperature is 25 °C for most inserts, and may be higher (up to 37 °C) for others. The incubation time should be optimised by the end user, and the following recommendations may be used as a guide:

• 0.1-1 kb: 5-10 mins
• 1-2 kb: 10-15 mins
• 2-3 kb: 15-20 mins
• ≥3 kb: 20-30 mins.

Note: If the insert is gel purified, use the maximum incubation time of 20-30 minutes.

#### Transformation

- 1. Add the ligated products to 50 μl of T1 Phage Resistant Chemically Competent Cells and mix gently. Do not mix by pipetting up and down.
- 2. Place on ice for 20-30 minutes.
- 3. Heat shock the cells at 42°C for 30 seconds, then immediately place the tube on ice for 2 minutes.
- 4. Add 250 μl of room temperature SOC or LB Medium, then incubate in a shaking incubator (200 rpm) at 37°C for 1 hour.
- 5. Mix 8 µl of 500 mM IPTG and 40 µl of 20 mg/ml X-gal, and spread evenly on a selective LB plate. Incubate at 37 °C for 30 minutes.
- 6. Inoculate a pre-warmed culture plate with 200  $\mu$ I of the transformants.

#### Identification of positive clones

- 1. Transfer 5-10 white or light-blue colonies into 10 µl of Nuclease-free water and vortex.
- 2. Use 1 µl of the mixture as the template for 25 µl PCR using M13 forward and M13 reverse primers.
- 3. Carry out PCR using the following reaction conditions:

| Number of Cycles | Temperature | Time per Cycle                          |
|------------------|-------------|---|
| 1 cycle          | 94 °C       | 10 min                                  |
|                  | 94 °C       | 30 seconds                              |
| 30 cycles        | 55 °C       | 30 seconds                              |
|                  | 72 °C       | Dependent on insert size and PCR Enzyme |
| 1 cycle          | 72 °C       | 5-10 min                                |

Note: PCR product size from vector self-ligation is 199 bp.

## Sequencing analysis

Positive clones should be inoculated in Ampicillin or Kanamycin selective LB liquid media, then incubate in a shaking incubator (200 rpm) at 37°C for 6 hours. Isolate plasmid DNA for restriction enzyme digestion and DNA sequencing. Sequencing should be performed using M13 forward, M13 Reverse and T7 promoter.

#### PCR of control insert (700 bp)

Ligate 1 µl of control insert with 1 µl vector, and carry out PCR according to the following conditions:

### Required Components

| Component                  | Volume   | Final concentration |
|----------------------------|----------|---------------------|
| Control template (5 ng/µl) | 1 µl     | 0.1 ng/μl           |
| Control Primers (10 µM)    | 1 µl     | 0.2 μΜ              |
| 2X PCR SuperMix            | 25 µl    | 1X                  |
| Nuclease-free water        | Variable | -                   |
| Total volume               | 50 µl    | -                   |
|                            |          |                     |

# PCR Conditions

| Number of Cycles | Temperature | Time per Cycle |
|------------------|-------------|----------------|
| 1 cycle          | 94 °C       | 2-5 min        |
|                  | 94 °C       | 30 seconds     |
| 30 cycles        | 50-60 °C    | 30 seconds     |
|                  | 72 °C       | 1 min          |
| 1 cycle          | 72 °C       | 10 min         |

#### Notes

- The PCR Enzyme should be a *Tag* DNA Polymerase.
- Primers must not be phosphorylated.
- 5-10 min post extension step is required. After amplification, agarose gel electrophoresis is recommended to validate the quality and quantity of the PCR products.